




The immune system and its never-ending battle against tumor cells: recognition, elimination, and evasion of the immune response

El sistema inmune y su interminable batalla contra las células tumorales: reconocimiento, eliminación y evasión de la respuesta inmune

Güitrón-Aviña, M. S,¹ , Ortiz-Lazareno, P. C,² , Solorzano-Ibarra, F. 1,^{3*} 

¹ Instituto de Investigación en Enfermedades Crónico-Degenerativas, Departamento de Biología Molecular y Genómica, Centro Universitario de Ciencias de la Salud, Universidad de Guadalajara, Guadalajara, Jalisco, México.

² División de Inmunología, Centro de Investigación Biomédica de Occidente, Instituto Mexicano del Seguro Social, Guadalajara, Jalisco, México.

³ Estancias posdoctorales por México.



Please cite this article as/Como citar este artículo: Güitrón-Aviña, M. S., Ortiz-Lazareno, P. C., Solorzano-Ibarra, F. (2024). The immune system and its never-ending battle against tumor cells. *Revista Bio Ciencias*, 11, e1614. <https://doi.org/10.15741/revbio.11.e1614>

Article Info/Información del artículo

Received/Recibido: December 05^h 2024.

Accepted/Aceptado: May 29^h 2024.

Available on line/Publicado: June 19^h 2024.

ABSTRACT

Cancer is one of the main causes of death in the world; for this reason, current research focuses on evaluating the mechanisms involved in the development of this disease. Over the years, it has been shown that the immune response (both innate and adaptive) plays a fundamental role in identifying and eliminating tumor cells. However, tumors can evade immune recognition and continue proliferating. In this review, we explain current aspects of the mechanisms used by the immune system to eliminate tumor cells; we also provide a review of the mechanisms mediated by the tumor to evade the immune response.

KEY WORDS: Immune response, cancer, innate immunity, adaptive immunity, evasion mechanisms.

*Corresponding Author:

Fabiola Solorzano-Ibarra. Investigadora postdoctoral, Instituto de Investigación en Enfermedades Crónico-Degenerativa. Departamento de Fisiología. Centro Universitario de Ciencias de la Salud. Sierra Mojada, 950, colonia Independencia. C.P 44340, Guadalajara, Jalisco, México. Teléfono: (33) 1058 5200. E-mail fabiolasolorzanoibarra@gmail.com

RESUMEN

El cáncer es una de las principales causas de muerte en el mundo, por ello las investigaciones actuales se centran en evaluar los mecanismos implicados en el desarrollo de esta enfermedad. A lo largo de los años se ha demostrado que la respuesta inmune (tanto innata como adaptativa) juega un papel fundamental en la identificación y eliminación de células tumorales. Sin embargo, los tumores pueden evadir el reconocimiento inmunológico y seguir proliferando. En esta revisión explicamos aspectos actuales sobre los mecanismos usados por el sistema inmune en humanos para eliminar las células tumorales, también hacemos una revisión de los mecanismos mediados por parte del tumor para evadir la respuesta inmune.

PALABRAS CLAVE: Respuesta inmune, cáncer, inmunidad innata, inmunidad adaptativa, mecanismos de evasión.

Introduction

Cancer is one of the main causes of death in the world. In 2020, ten million people died from this disease (Sung *et al.*, 2021). The etiology of cancer is complex and depends on a diversity of factors that include aging, lifestyle habits, and genetic aspects, which together can induce permanent (but not lethal) errors in the DNA (Deoxyribonucleic Acid) of the cell, initiating the process of tumor transformation (Wu *et al.*, 2016). Tumor cells acquire diverse biological capabilities that help evade cell cycle regulation and allow them to continue proliferating. These capabilities include proliferative signaling, evasion of growth suppressor genes, resistance to cell death, replicative immortality, angiogenesis induction, invasion, metastasis, reprogramming of cellular metabolism, and evasion of destruction by the immune response (Hanahan, 2022).

Tumors are more than masses of transformed cells; they are also composed of other cellular components, including the infiltration of immune cells, that eliminate transformed cells to prevent cancer development (Burnet, 1964). However, tumors have acquired the ability to evade the immune response since they reduce the expression of tumor antigens and MHC (Main Histocompatibility Complex) molecules. Tumors also secrete cytokines that suppress the function of immune cells (Dunn *et al.*, 2004^a; Dunn *et al.*, 2004^b). In this article, we explain current aspects of the mechanisms of tumor elimination mediated by the immune system. On the other hand, we also review the escape mechanisms that the tumor uses to evade the immune response.

Carcinogenesis process

Carcinogenesis is the process of malignant cell transformation, in which alterations in the DNA and failures in the signaling pathways that control and regulate cell proliferation are observed. Subsequently, cells continue to proliferate, accumulating more mutations and genetic changes. This allows tumor cells to acquire distinctive capabilities known as the hallmarks of cancer, which were first proposed in 2000 and expanded in 2011 (Hanahan & Weinberg, 2000). These hallmarks include the ability to sustain proliferative signaling, evade growth suppressors, resist cell death, enable replicative immortality, induce angiogenesis, activate invasion and metastasis, metabolic reprogramming, induce inflammation, genomic instability, and evasion of immune destruction. Additional emerging characteristics were recently included: unlocking phenotypic plasticity, and non-mutational epigenetic reprogramming. There are also factors, such as polymorphic variability in the microbiomes of the tumor microenvironment, as well as senescent cells, that can modify the capabilities acquired by tumor cells and impact cancer phenotypes (Hanahan, 2022). The characteristics acquired by transformed cells allow them to detach from the tissue and travel through the bloodstream to a secondary organ, where the tumor cell adheres again and begins to proliferate, generating new blood vessels and allowing cancer cells to spread to other parts of the body. This entire process, known as metastasis, is observed in patients in advanced stages of the disease (Lambert *et al.*, 2017).

During cancer development, the damage in a cell caused by genetic aspects and lifestyle is required. Those aspects are considered factors that increase the risk of suffering cancer and are classified as non-modifiable and modifiable (Cogliano *et al.*, 2011; Wu *et al.*, 2018). Non-modifiable risk factors include mutations that occur spontaneously or are inherited in the genome. Spontaneous mutations arise due to random errors in DNA replication during the cell cycle and are acquired at some point in the life of any person (Tomasetti & Vogelstein, 2015). There are also mutations inherited from the parents that are not modifiable and are associated with the development of inherited cancers, which are rare and represent 5 to 10% of all cancers (Fearon, 1997; Lichtenstein *et al.*, 2000; Jahn *et al.*, 2022; Garutti *et al.*, 2023).

On the other hand, modifiable risk factors are related to lifestyle and the environment (diet, physical activity, toxic compounds, sun exposure, and, infections by microbes or viruses) (Wu *et al.*, 2018). Those factors are considered modifiable because a healthy lifestyle can reduce the risk of cancer. For example, exposure to ultraviolet radiation is the main risk factor for the development of skin cancer, since it is considered a carcinogen that induces DNA mutations and suppresses the antitumor immune response (Neale *et al.*, 2023). It has been reported that people who experience more than 5 episodes of severe sunburn have a 2 times greater risk of developing melanoma, therefore, avoiding exposure to the sun for long periods and using sun protection can reduce the possibility of suffering from this type of cancer (White & Lee, 1994). Infections caused by oncogenic microbes or viruses have been related to the formation of tumors, an example is the persistent infection by the Human Papillomavirus (mainly by genotypes 16 and 18), which is associated with approximately 95-99.7% of the cases of cervical cancer (Bosch *et al.*, 1995; Kusakabe *et al.*, 2023).

Role of the immune system in the development of cancer

Tumors are more than clusters of transformed cells; they are also composed of the extra cellular matrix (chemokines, inflammatory cytokines, integrins, matrix metalloproteinases, and other secreted molecules), stromal cells (endothelial cells and fibroblasts), immune cells and other resident cells of the tissue, which together constitute a complex and dynamic ecosystem known as the tumor microenvironment, in which there is an interaction between all its components (Dvorak, 1986; Clemente *et al.*, 1996; Chen *et al.*, 2017; Ho *et al.*, 2022). Depending on the tumor stage, the organ where it formed, the characteristics of the tumor cells and the patient, the components of the tumor microenvironment, and their functional status may vary (de Visser & Joyce, 2023). Immune cells in the tumor microenvironment can be located differently, which affects the communication with the tumor, for example, there are infiltrated tumors in which immune cells are homogeneously distributed throughout the tumor. In other tumors, the immune components are distributed in the periphery without infiltrating the tumor; these are considered immune-excluded tumors. While so-called immune-silent tumors that lack immune cells can also be observed (Anderson & Simon, 2020). Therefore, the interaction between the immune system and the tumor is complex and can have a dual effect, that is, it can suppress tumor development or promote tumorigenesis (Dunn *et al.*, 2002; Schreiber & Smyth, 2011; Anderson & Simon, 2020).

The role of the immune response in cancer has been a focus of research since the 1950s; In 1957, Sir Frank Macfarlane Burnet proposed the concept of “immunovigilance,” which explains the ability of the immune system to recognize and eliminate tumor cells. This theory was refuted and resurfaced several times (Burnet 1970). Subsequently, with technological advances, studies were carried out in murine models that made it possible to demonstrate that an immune response in which there is a lack of cells such as lymphocytes (T cells, B cells, and NK cells) or cytokines (a particular case of Interleukin 12 and Interferon- γ) is associated with the development of spontaneous tumors, induced by chemical or infectious agents such as viruses (Nishizuka *et al.*, 1965; Burstein & Law, 1971; Sanford *et al.*, 1973; Stutman, 1975; Dighe *et al.*, 1994; Kaplan *et al.*, 1998; Smyth *et al.*, 2000^a; Smyth *et al.*, 2000^b; Girardi *et al.*, 2001; Shankaran *et al.*, 2001; Street *et al.*, 2002). This hypothesis was later evaluated in humans and it was observed that patients suffering from autoimmune diseases or being treated with immunosuppressive drugs have a higher risk of developing cancer (Knuth *et al.*, 1984; van der Bruggen *et al.*, 1991; Sahin *et al.*, 1995). Currently, part of the new therapies against malignant tumors aim to modulate the immune response to eliminate transformed cells effectively (Korman *et al.*, 2006). The most promising therapies include immune checkpoint blockade (examples via PD-1/PD-L1, and CTLA-4), administration of cytokines (IL-2, Interleukin-2 and GM-CSF, Colony-Stimulating Factor of Granulocytes and Macrophages), adoptive TILs (Tumorinfiltrating Lymphocytes), and CAR-Ts (Genetically Engineered T cells expressing specific T cell Receptors, TCRs, or Chimeric Antigen Receptors CARs). (Hamdan & Cerullo, 2023). Selection of the best therapeutic option for a patient depends largely on the clinical stage, tumor type, tumor microenvironment, and the response to conventional treatments (chemotherapy or radiotherapy).

Recognition and elimination of tumor cells by the immune response

The immune system comprises innate immune cells (macrophages, neutrophils, dendritic cells, NK cells, among others) and adaptive immune cells (T lymphocytes and B lymphocytes). The innate immune system provides rapid defense mechanisms against tumors. In contrast, the adaptive immune system responds later, more specifically, which requires time and leads to the generation of immunological memory. Both types of responses are coordinated and work together to prevent the development of cancer (See Figure 1) (Hellstrom *et al.*, 1968; Herberman & Holden, 1978; Boon *et al.*, 1995; Anichini *et al.*, 1999, Girardi *et al.*, 2001; Schlosser *et al.*, 2019).

Recognition of Tumor Cells

During the process of malignant transformation, changes occur in the expression of antigens that differentiate tumor cells from healthy cells, inducing the recognition by the immune system (Robbins *et al.*, 2013). Different types of antigens can be recognized by immune cells. Molecules such as phosphatidylserines or calreticulins are expressed in the tumor cells and act as pro-phagocytic signals (Feng *et al.*, 2015; Vallabhapurapu *et al.*, 2015).

All the cells that undergo a stress process can suffer a type of death “called immunological death,” in which the cell death is accompanied by the secretion of molecules called Damage-Associated Molecular Patterns (DAMPs). (Galluzzi *et al.*, 2020). These molecules can activate the immune system and function as warning molecules for the innate immune cells. DAMPs are identified by PRRs (Pattern Recognition Receptors) expressed in cells such as macrophages, neutrophils, and dendritic cells (Matzinger, 1994; Bluwstein *et al.*, 2013; Gong *et al.*, 2020).

Tumor cells exhibit high expression of heat shock proteins that can be captured by dendritic cells, processed, and presented to CD8 T cells. An example is the heat shock protein Hsp70 (Tumor-derived Heat Shock Protein (HSP)70) (Noessner *et al.*, 2002). On the other hand, it has been reported that HSP70 also mediates the NK cells activation and promotes cytotoxicity *in vitro* (Gross *et al.*, 2008).

Due to the number of mutations in the genome of tumor cells, the expression of proteins with a low expression in healthy cells increases. An example is the protein NY-ESO-1 (New York Esophageal Squamous Cell Carcinoma 1) expressed in germ or placental cells, and overexpressed in several types of cancer (melanoma, sarcoma, and some types of lung cancer, breast, ovary, among others) (Jungbluth *et al.*, 2001; Satie *et al.*, 2002; Woloszynska-Read *et al.*, 2008, Thomas *et al.*, 2018). Tumor cells also produce new incomplete, non-functional, or defective proteins that are identified by the immune system, these are known as neoantigens and are produced by tumor cells due to genomic mutations, aberrant transcriptomic variants, and post-translational modifications (Yarchoan *et al.*, 2017; Smith *et al.*, 2019). Neoantigens are widely studied in the design of personalized vaccines (peptide, nucleic acid, and dendritic cell vaccines). The most immunogenic neoantigens are used as targets in adoptive cell therapies that use autologous TILs (Tumor-Infiltrating Lymphocytes) expanded *in vitro* without genetic modifications and genetically

modified immune cells TCRs (T-Cell Receptor) or CARs (Chimeric Antigen Receptor) (Liu *et al.*, 2019; Yamamoto & Restifo, 2019; Pajens *et al.*, 2021). Finally, therapies based on antibodies against neoantigens have also been developed for various purposes. For example, antibodies conjugated with drugs directed against a neoantigen are more effective when administered to the tumor site.

Phagocytic cells

Phagocytic cells (monocyte/macrophage, dendritic cell, neutrophil, mast cell, and B cells) are responsible for capturing apoptotic cells and degrading them (Feng *et al.*, 2019). For example, macrophages recognize DAMPs in apoptotic transformed cells and engulf them, thus eliminating tumor cell debris. Tumor-Associated Macrophages (TAMs) are composed of two main populations with distinct phenotypes: M1 (classically activated macrophages) and M2 (alternatively activated macrophages) (Mills *et al.*, 2000). The polarization to each subtype depends on different stimuli (Shapouri-Moghaddam *et al.*, 2018) Stimulation with IFN- γ and/or bacterial lipopolysaccharides drives macrophage polarization to an M1, which has a pro-inflammatory and anti-tumor role due to the high expression of MHC molecules (which increase the presentation of tumor antigens), nitric oxide, reactive oxygen, and nitrogen species. Furthermore, M1 macrophages secrete a great diversity of cytokines such as TNF- α , IL-1 α , IL-1 β , IL-6, IL-12, IL-18, and IL-23, which modulate the immune response against the tumor (Duan & Luo, 2021). On the other hand, polarization to an M2 (anti-inflammatory) is associated with cytokines such as IL-4 and IL-13, in addition, it participates in tissue remodeling regeneration, and wound healing through the production of arginase/ornithine, EGF, VEGF, and TGF- β . In cancer, M2 promotes tumor growth by downregulating the immune response (Mantovani *et al.*, 2013; Mills & Ley, 2014; Duan & Luo, 2021). TAMs are mostly macrophages of the M2 type, for this reason the presence of these cells is associated with tumor progression, poor prognosis, and resistance to immunotherapy (Gao & Wang, 2022).

Another type of phagocytic cell are dendritic cells, which mainly capture tumor antigens, phagocytose them, degrade them, and assemble them into MHC molecules. These cells then travel to the lymph nodes that drain the tumor and present the antigens on the MHC molecules to CD4 T cells (known as helpers for their function in coordinating immune responses) and CD8 T cells (with cytotoxic functions that help destroy tumor cells). This process of antigen presentation allows the activation of T cells and the expansion of clones of antigen-specific CD4 or CD8 T cells (Nussenzweig *et al.*, 1980; Albert & Bhardwaj, 1998; Steinman, 2012). Dendritic cells can be classified as conventional or classical (cDC) and plasmacytoid (pDC). The cDCs are also classified into cDC1 and cDC2 (Del Prete *et al.*, 2023). cDC1 mainly specializes in the processing and cross-presenting of antigens derived from necrotic and apoptotic tumor cells associated with MHC I molecules to CD8 T cells (See *et al.*, 2017). There is a correlation between the proportion of cDC1 in the tumor with survival and response to treatment in cancer patients. Regarding cDC2, these cells present antigens associated with MHC II molecules to CD4 T cells to promote the polarization of T helper cells to a Th1, Th2, and Th17 profile, with different functions in cancer (Plesca *et al.*, 2022). A higher frequency of cDC2s correlates with higher CD4 T cell infiltration in melanoma (Binnewies *et al.*, 2019). Regarding plasmacytoid dendritic cells, they are the main producers of type I interferons (IFN), cytokines that play an important role in cancer by modulating

the immune response to eliminate tumor cells (Reizis, 2019). Furthermore, pDCs contribute to the presentation of antigens to CD4 T cells via MHC II molecules (Siegal *et al.*, 1999).

Neutrophils are a population of phagocytic cells that represent an important part of the immune cells in the tumor microenvironment of various types of cancer, including colorectal cancer, cervical cancer, and non-small cell lung cancer, among others (Rao *et al.*, 2012; Carus *et al.*, 2013; Rakaee *et al.*, 2016). It is reported that the production of Granulocyte Colony-Stimulating Factor (G-CSF) by tumor cells favors the release of neutrophils into the circulation, increasing their proportions in the blood (Jablonska *et al.*, 2017; Furumaya *et al.*, 2020). Furthermore, tumor cells can secrete IL-8 and attract neutrophils and other myeloid cells to the tumor microenvironment (Gonzalez-Aparicio & Alfaro, 2020). Tumor-Associated neutrophils can induce direct cytotoxicity and inhibition of metastasis; however, they represent a heterogeneous population of cells that may have both anti- and pro-tumor roles in cancer (Shaul & Fridlender, 2019). In murine models, neutrophils are classified as antitumor neutrophils (N1) or tumor-promoting neutrophils (N2) (Fridlender *et al.*, 2009). Three populations of neutrophils classified according to their densities after isolation by centrifugation have been identified in the peripheral blood of cancer patients: High-Density mature neutrophils (HDN), Low-Density mature neutrophils (mature LDN) and Low-Density immature neutrophils (immature LDN) (Sagiv *et al.*, 2015; Shaul & Fridlender, 2019). HDNs have an antitumor effect similar to N1, while mature LDNs or immature LDNs have a protumor effect (Sagiv *et al.*, 2015). Mast cells are another population of cells that have recently been studied in the context of cancer because of their dual role (pro-tumor or anti-tumor) depending on the type of tumor and location in the tumor microenvironment (Varricchi *et al.*, 2017). For example, it has been observed that they have a pro-tumoral role in different types of cancer like gastric, thyroid, pancreatic, bladder, Merkel cell carcinoma, Hodgkin, and non-Hodgkin lymphoma. Mast cells can even be associated with poor prognosis (Agata *et al.*, 1996; Yano *et al.*, 1999; Beer *et al.*, 2008; Melillo *et al.*, 2010; Rabenhorst *et al.*, 2012; Andersen *et al.*, 2016; Rao *et al.*, 2016). On the contrary, in breast cancer, mast cells have antitumor functions (Dabiri *et al.*, 2004). The antitumor activity of mast cells is mediated by the release of ROS and TNF- α , which have a cytotoxic effect, and the release of heparin and IL-9, which inhibit tumor growth. Furthermore, histamine release contributes to the maturation of dendritic cells (Varricchi *et al.*, 2017).

B cells and the role of antibodies in cancer

B lymphocytes are important in cancer as they are antigen-presenting cells to CD4 T cells and antibody-producing cells (Chen & Jensen 2008; DiLillo *et al.*, 2010). For a B cell to be activated, it must first recognize the tumor antigen through its BCR; this antigen is then processed and presented to CD4 T cells. This interaction provides the costimulatory signals necessary to complete B cell activation that allows their clonal expansion, generating a population of identical B cells with specificity for the same antigen and capable of producing antibodies. Some activated B cells differentiate into plasma cells, the main producers of antibodies (LeBien & Tedder, 2008).

Antibodies are soluble molecules that recognize and bind to an antigen through a highly specific and complementary chemical interaction between the antigen-binding site molecules (present on the antibody) and the chemical structure of the antigen.

The functions of antibodies in cancer are diverse. By binding to tumor cells, they allow macrophages and dendritic cells to capture tumor antigens since they induce phagocytosis (Anderson *et al.*, 1990; Ravetch & Bolland, 2001; Tan & Long, 2022). They also promote antibody-dependent cellular cytotoxicity (ADCC), a process in which cytotoxic cells such as NK cells (Natural Killer cells) recognize antibodies bound to the tumor, leading to the release of molecules that induce tumor cell death (Titus *et al.*, 1987; Sondel & Hank, 2001; Bruhns, 2012).

Antibodies bound to tumor cells also promote the recognition by the complement system, a group of proteins that form a complex attack on the membrane of tumor cells that leads to cell lysis and destruction (Kolev *et al.*, 2022). However, the function of antibodies is not always beneficial in cancer since antibodies can bind to various soluble antigens released by tumor cells, which favors the formation of circulating immune complexes that have negative implications (Gunderson & Coussens, 2013; Tan & Long, 2022). In a murine model of squamous cell carcinoma, the accumulation of immune complexes in the stroma of the tumor tissue increases the recruitment of protumoral myeloid cells (Andreu *et al.*, 2010).

Cytotoxic lymphocytes and CD4 T cells

In the immune system, we can find two types of cytotoxic cells: Natural Killer (NK) cells and CD8 T cells (counterpart of the adaptive immunity) (West *et al.*, 1977). Both cells destroy transformed cells through the secretion of granules with granzymes and perforins, which induce the death of the target cell (Sykulev *et al.*, 1996; Barry & Bleackley, 2002).

NK cells belong to the group of innate lymphoid cells and are characterized by the expression of the transcription factor Eomesodermin (EOMES) (Cherrier *et al.*, 2018). Unlike CD8 T cells that express antigen-specific clonotypic receptors, NK cells are equipped with an arsenal of germline-encoded activating or inhibitory receptors, which recognize their cognate ligands on healthy, infected, and transformed cells (Karre, 1997; Diefenbach & Raulet, 1999; Bryceson *et al.*, 2006; Lanier, 2008). Tumor cells can express molecules recognized by activating receptors, such as MICA (activating ligand for the NKG2D receptor) overexpressed on cells in stress situations. The recognition of MICA by NKG2D induces the activation in the NK cell and the secretion of cytotoxic granules. NK cells are also activated in the absence of MHC I molecules (a common mechanism developed by tumors to evade the recognition by CD8 T cells) (Karre, 1997). A great diversity of activation receptors is expressed by the NK cells; among the most studied are the NCRs family members (Natural Cytotoxicity Receptors, such as NKp30, NKp44, and NKp46) and the members of the NKG2 family (NKG2C and NKG2D). The main inhibitory receptors include members of the KIR (Killer Immunoglobulin-like Receptors) family and NKG2A (NKG2 family member) (Nersesian *et al.*, 2023). The expression and co-expression of the various receptors are associated with NK cell populations with a different maturation status and functions. In cancer, Tumor-Infiltrating NK cells are associated with a good disease prognosis. Therefore, the modulation of the response mediated by NK cells is an important therapeutic target in cancer. The main strategies include the induction of NK cell activation by administering monoclonal antibodies to increase ADCC, BiKEs (Bispecific Killer Cell Engager), *ex vivo* expanded NK cells, and CAR-NK (Liu *et al.*, 2020; Dixon *et al.*, 2021).

Regarding CD8 T cells, they recognize specific tumor antigens presented on MHC I molecules. This presentation occurs in the lymph nodes close to the tumor, and to complete the activation, other costimulatory signals (CD28 or NKG2D) and cytokines produced by the dendritic cell are required. This process induces the proliferation, clonal expansion, and differentiation of naïve CD8 T cells into effector cells, where subsequently, CD8 T cell clones migrate to the tumor microenvironment, recognize the antigen on the tumor cells, and release their cytotoxic granules to eliminate tumor cells. CD8 T cells also produce cytokines such as IFN- γ and TNF- α (Koh *et al.*, 2023).

Other cells have cytotoxic functions in cancer; for example, the antitumor function of CD4 T cells depends on their ability to directly activate CD8 T cells and the activation of macrophages and NK cells. On the other hand, a population of cytotoxic CD4 T cells capable of secreting granzymes in a similar way to CD8 T cells is observed in cancer. Another example is the case of B lymphocytes that can directly attack tumor cells using granzyme B and the TRAIL ligand (TNF Related Apoptosis-Inducing Ligand) that will induce tumor cell death by interacting with its cognate molecule (Van-Leeuwen *et al.*, 2004; Quezada *et al.*, 2010; Janjic *et al.*, 2022). Other cells with a cytotoxic function in the tumor microenvironment are macrophages that release reactive oxygen species and reactive nitrogen species that contribute to the destruction of cancer cells.

Mechanisms of evasion of the immune response in cancer

Although the immune system can eliminate tumor cells, it can also edit the tumor by eliminating the most immunogenic clones. Therefore, the less immunogenic tumor cells proliferate without control (Schreiber & Smyth, 2011). This idea is the basis of the cancer immunoediting hypothesis, which consists of three distinct phases: elimination, equilibrium, and escape (Dunn *et al.*; 2002; Dunn *et al.*, 2004a; Dunn *et al.*, 2004b; Vesely *et al.*, 2011). The elimination phase includes the stage in which the immune system eliminates the most immunogenic tumor cells, through the mechanisms described in the previous sections. In the equilibrium phase, the cell variants that survived the elimination by the immune system move to this stage, where adaptive immune cells control the growth of tumor cells and maintain them in a functional state of latency for decades (Aguirre-Ghiso, 2007). While in the escape phase, tumor cells that evaded the elimination phase, because they acquired mechanisms to evade the immune response, begin to proliferate rapidly, forming detectable tumors and advanced stages of the disease (Smyth *et al.*, 2006).

A mechanism used by tumors to evade the immune response is the loss of the expression of tumor antigens, which prevents tumor cells from being identified by the immune system. For example, tumors reduce the expression of MHC I molecules, avoiding detection by T cells (as previously described) (Dhatchinamoorthy *et al.*, 2021). They also secrete metalloproteases that cleave the activating ligands for NK cells. A clear example is the proteolytic cleavage of MICA, in this way tumor cells escape the elimination mediated by NK cells (Deng *et al.*, 2015; Xing & Ferrari de Andrade, 2020). Some tumors secrete cytokines that suppress the immune system and create an immunosuppressive tumor microenvironment for example IL-10 and TGF- β (Transforming Growth Factor-Beta). One of the functions of IL-10 is downregulating the expression of MHC II molecules in antigen-presenting cells, while TGF- β inhibits the activation of T cells and NK cells (Fischer *et al.*, 1992; Lazar-Molnar *et al.*, 2000). The immunosuppressive tumor microenvironment

induces the expansion of regulatory T or B cells (variants of CD4 T cells or B cells with the function of inhibiting the activation of other immune cells and their functions) (Mauri & Bosma, 2012; Togashi *et al.*, 2019). Another mechanism of evasion of cancer cells is the expression of molecules such as PD-L1 (present in the tumor) that when interacting with its receptor PD-1 (expressed in B lymphocytes, CD8 T cells, CD4 T cells, NK cells, among others) inhibits the function of immune cells (Agata *et al.*, 1996; Liu *et al.*, 2021). The constant stimulation with this type of inhibitory molecules, together with a suppressive microenvironment, hypoxia, and the presence of tumor antigens, induces a process in CD8 T cells known as exhaustion, in which T cells begin to gradually lose their functions including their cytotoxic effect (Wherry, 2011).

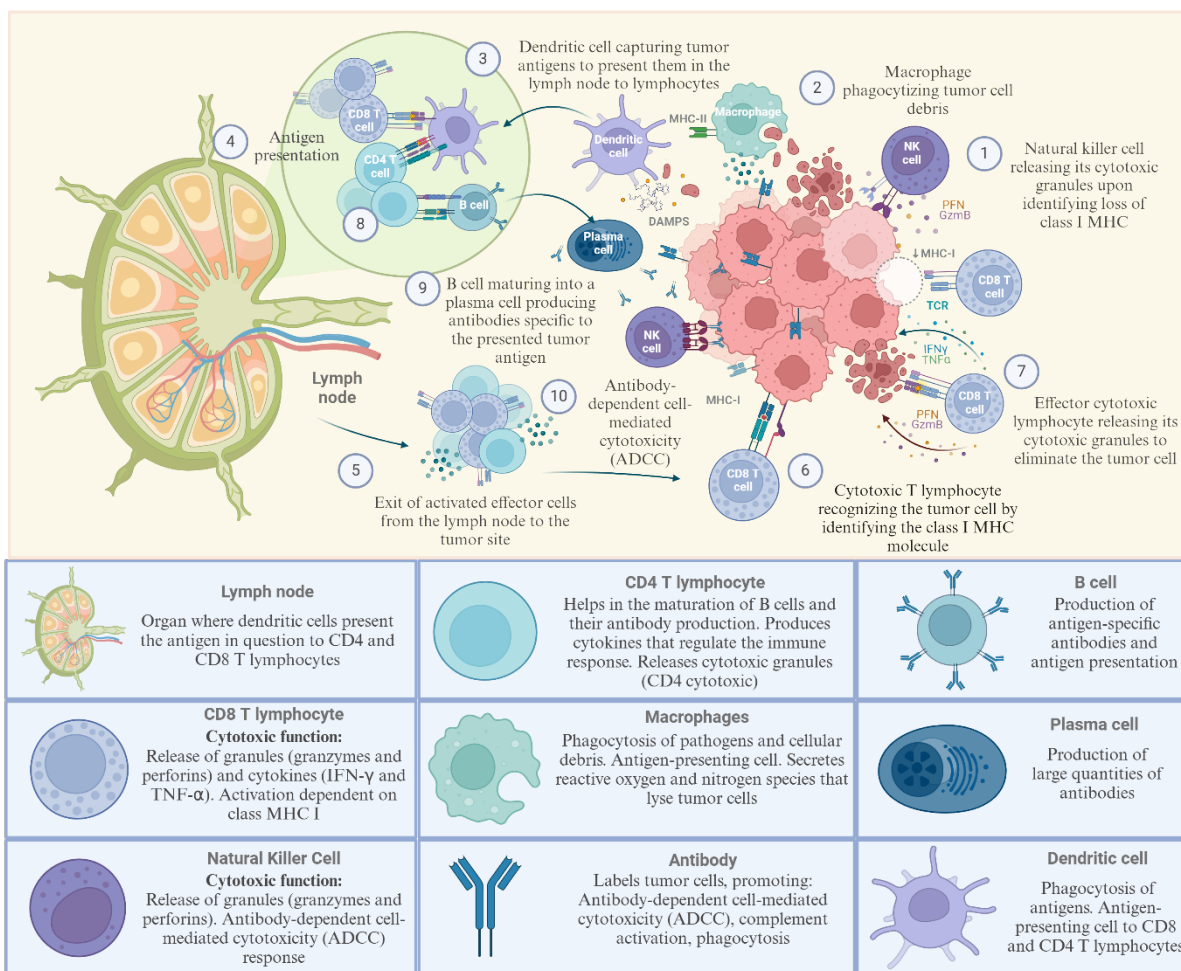


Figure 1. Immune response against cancer.

Different cells and molecules that comprise the immune system, such as macrophages, dendritic cells, NK cells, T cells (CD4 and CD8), and B cells, work together to eliminate tumor cells. Transformed cells express

molecules called antigens that induce an immune response. (1) Cells of the innate immune system, such as NK cells, release their cytotoxic granules (with granzymes and perforins) upon the recognition of tumor cells. (2) Macrophages identify antigens and phagocytose apoptotic tumor cells and present them in MHC class I and II molecules. (3) Similarly, dendritic cells take up these tumor antigens to present them to CD4 and CD8 T cells in the lymph nodes near the tumor. This process is the union between the innate and adaptive immune systems. (4) Antigen presentation occurs after the TCR recognizes the antigen bound in MHC I or MHC II molecules. (5) This generates the clonal expansion and migration of effector T cells (CD4 or CD8) to the tumor microenvironment. (6) Once in the tumor, CD8 T cells recognize the tumor cell by identifying the MHC I molecule. (7) In this way, the effector lymphocytes are activated and release their cytotoxic granules (with granzymes, perforins) and cytokines (IFN- γ , Interferon- γ , and TNF Tumor Necrosis Factor) to eliminate the tumor cells. (8) B cells identify antigens on tumor cells, phagocytose them, and present them to CD4 T cells. This interaction induces the activation of B cells. (9) Cooperation with CD4 cells helps the maturation of B cells into plasma cells, which produce large amounts of antibodies. (10) Finally, antibodies produced by these cells bind to tumor antigens against which they were generated and allow phagocytosis by phagocytic cells or antibody-mediated cytotoxicity by NK cells.

Image created in Biorender.

Conclusions

The immune system comprises cells and molecules that can prevent cancer development by identifying and destroying tumor cells. However, tumors acquire mutations that induce the secretion of cytokines or expression of immunomodulatory molecules that tumors use in advantage to continue proliferating without being eliminated. Therefore, cancer research has focused on studying the tumor immune evasion mechanisms to understand the interaction between immune cells and transformed cells. This knowledge helps to develop immunotherapies that modify the immune response by reinvigorating or changing the phenotype of immune cells. Together, it helps to have a specific recognition of the tumor cells and a more effective elimination. New research should focus on evaluating changes in the tumor microenvironment regarding the expression profile of inhibition and activation receptors with therapeutic potential and cytokines that allow the activation of cytotoxic cells or polarization to an anti-tumor profile.

Author contribution

Conceptualization of the work: S.I.F., O.L.P.C.; Writing and manuscript preparation: G.A.M.S., S.I.F.; Writing, review, and editing: G.A.M.S., S.I.F., O.L.P.C.; Figure creation and editing: : G.A.M.S., S.I.F., O.L.P.C..

“All authors of this manuscript have read and accepted the published version”.

Acknowledgments

Gratitude for the support provided by the Postdoctoral Stays Program for Mexico CONACYT.

Interest conflict

“The authors declare no conflict of interest”.

References

- Agata, Y., Kawasaki, A., Nishimura, H., Ishida, Y., Tsubata, T., Yagita, H., & Honjo, T. (1996). Expression of the PD-1 antigen on the surface of stimulated mouse T and B lymphocytes. *International Immunology*, 8(5), 765-772. <https://doi.org/10.1093/intimm/8.5.765>
- Aguirre-Ghiso, J. A. (2007). Models, mechanisms and clinical evidence for cancer dormancy. *Nature Reviews Cancer*, 7(11), 834-846. <https://doi.org/10.1038/nrc2256>
- Albert, M. L., Sauter, B., & Bhardwaj, N. (1998). Dendritic cells acquire antigen from apoptotic cells and induce class I-restricted CTLs. *Nature*, 392(6671), 86-89. <https://doi.org/10.1038/32183>
- Andersen, M. D., Kamper, P., Nielsen, P. S., Bendix, K., Riber-Hansen, R., Steiniche, T., Hamilton-Dutoit, S., Clausen, M., & d'Amore, F. (2016). Tumour-associated mast cells in classical Hodgkin's lymphoma: correlation with histological subtype, other tumour-infiltrating inflammatory cell subsets and outcome. *European Journal Haematology*, 96(3), 252-259. <https://doi.org/10.1111/ejh.12583>
- Anderson, C. L., Shen, L., Eicher, D. M., Wewers, M. D., & Gill, J. K. (1990). Phagocytosis mediated by three distinct Fc gamma receptor classes on human leukocytes. *Journal of Experimental Medicine*, 171(4), 1333-1345. <https://doi.org/10.1084/jem.171.4.1333>
- Anderson, N. M., & Simon, M. C. (2020). The tumor microenvironment. *Current Biology*, 30(16), R921-R925. <https://doi.org/10.1016/j.cub.2020.06.081>
- Andreu, P., Johansson, M., Affara, N. I., Pucci, F., Tan, T., Junankar, S., Korets, L., Lam, J., Tawfik, D., DeNardo, D. G., Naldini, L., de Visser, K. E., De Palma, M., & Coussens, L. M. (2010). FcRgamma activation regulates inflammation-associated squamous carcinogenesis. *Cancer Cell*, 17(2), 121-134. <https://doi.org/10.1016/j.ccr.2009.12.019>
- Anichini, A., Molla, A., Mortarini, R., Tragni, G., Bersani, I., Di Nicola, M., Gianni, A. M., Pilotti, S., Dunbar, R., Cerundolo, V., & Parmiani, G. (1999). An Expanded Peripheral T Cell Population to a Cytotoxic T Lymphocyte (CtI)-Defined, Melanocyte-Specific Antigen in Metastatic Melanoma Patients Impacts on Generation of Peptide-Specific Ctls but Does Not Overcome Tumor Escape from Immune Surveillance in Metastatic Lesions. *The Journal of Experimental Medicine*, 190, 651- 668. <https://doi.org/10.1084/jem.190.5.651>
- Barry, M., & Bleackley, R. C. (2002). Cytotoxic T lymphocytes: all roads lead to death. *Nature Reviews Immunology*, 2(6), 401-409. DOI: <https://doi.org/10.1038/nri819>
- Beer, T. W., Ng, L. B., & Murray, K. (2008). Mast cells have prognostic value in Merkel cell carcinoma. *The American Journal Dermatopathology*, 30(1), 27-30. <https://doi.org/10.1097/dad.0b013e31815c932a>
- Binnewies, M., Mujal, A. M., Pollack, J. L., Combes, A. J., Hardison, E. A., Barry, K. C., Tsui, J., Ruhland, M. K., Kersten, K., Abushawish, M. A., Spasic, M., Giurintano, J. P., Chan, V., Daud, A. I., Ha, P., Ye, C. J., Roberts, E. W., & Krummel, M. F. (2019). Unleashing Type-2 Dendritic Cells to Drive Protective Antitumor CD4(+) T Cell Immunity. *Cell*, 177(3), 556-571

- e516. <https://pubmed.ncbi.nlm.nih.gov/30955881/>
- Bluwstein, A., Kumar, N., Léger, K., Traenkle, J., Oostrum, J. V., Rehrauer, H., Baudis, M., & Hottiger, M. O. (2013). PKC signaling prevents irradiation-induced apoptosis of primary human fibroblasts. *Cell Death & Disease*, 4(2), e498-e498. <https://doi.org/10.1038/cddis.2013.15>
- Boon, T., Gajewski, T. F., & Coullie, V. (1995). From defined human tumor antigens to effective immunization? *Immunology Today*, 16(7), 334-336. [https://doi.org/10.1016/0167-5699\(95\)80149-9](https://doi.org/10.1016/0167-5699(95)80149-9)
- Bosch, F. X., Manos, M. M., Munoz, N., Sherman, M., Jansen, A. M., Peto, J., Schiffman, M. H., Moreno, V., Kurman, R., & Shah, K. V. (1995). Prevalence of human papillomavirus in cervical cancer: a worldwide perspective. International biological study on cervical cancer (IBSCC) Study Group. *Journal of the National Cancer Institute*, 87(11), 796-802. <https://doi.org/10.1093/jnci/87.11.796>
- Bruhns, P. (2012). Properties of mouse and human IgG receptors and their contribution to disease models. *Blood*, 119(24), 5640-5649. <https://doi.org/10.1182/blood-2012-01-380121>
- Bryceson, Y. T., March, M. E., Ljunggren, H.-G., & Long, E. O. (2006). Synergy among receptors on resting NK cells for the activation of natural cytotoxicity and cytokine secretion. *Blood*, 107(1), 159-166. <https://doi.org/10.1182/blood-2005-04-1351>
- Burnet, F. M. (1970). The concept of immunological surveillance. *Progrès de la recherche expérimentale des tumeurs*, 13, 1-27. <https://doi.org/10.1159/000386035>
- Burnet, M. (1964). Immunological Factors in the Process of Carcinogenesis. *British Medical Bulletin*, 20, 154-158. <https://doi.org/10.1093/oxfordjournals.bmb.a070310>
- Burstein, N. A., & Law, L. W. (1971). Neonatal thymectomy and non-viral mammary tumours in mice. *Nature*, 231(5303), 450-452. <https://doi.org/10.1038/231450a0>
- Carus, A., Ladekar, M., Hager, H., Nedergaard, B. S., & Donskov, F. (2013). Tumour-associated CD66b+ neutrophil count is an independent prognostic factor for recurrence in localised cervical cancer. *British Journal of Cancer*, 108(10), 2116-2122. <https://doi.org/10.1038/bjc.2013.167>
- Chen, X., Chen, J., Zhang, W., Sun, R., Liu, T., Zheng, Y., & Wu, Y. (2017). Prognostic value of diametrically polarized tumor-associated macrophages in multiple myeloma. *Oncotarget*, 8(68), 112685-112696. <https://doi.org/10.18632/oncotarget.22340>
- Chen, X., & Jensen, P. E. (2008). The role of B lymphocytes as antigen-presenting cells. *Archivum Immunologiae et Therapiae Experimentalis*, 56(2), 77-83. <https://doi.org/10.1007/s00005-008-0014-5>
- Cherrier, D. E., Serafini, N., & Di Santo, J. P. (2018). Innate Lymphoid Cell Development: A T Cell Perspective. *Immunity*, 48(6), 1091-1103. <https://doi.org/10.1016/j.immuni.2018.05.010>
- Clemente, C. G., Mihm Jr, M. C., F. A. C. P., Bufalino, R., Zurrida, S., Collini, P., & Cascinelli, N. (1996). Prognostic value of tumor infiltrating lymphocytes in the vertical growth phase of primary cutaneous melanoma. *Cancer*, 77(7), 1303-1310. [https://doi.org/10.1002/\(sici\)1097-0142\(19960401\)77:7%3C1303::aid-cnrc12%3E3.0.co;2-5](https://doi.org/10.1002/(sici)1097-0142(19960401)77:7%3C1303::aid-cnrc12%3E3.0.co;2-5)
- Cogliano, V. J., Baan, R., Straif, K., Grosse, Y., Lauby-Secretan, B., El Ghissassi, F., Bouvard, V., Benbrahim-Tallaa, L., Guha, N., Freeman, C., Galichet, L., & Wild, C. P. (2011). Preventable exposures associated with human cancers. *Journal of the National Cancer Institute*, 103(24), 1827-1839. <https://doi.org/10.1093/jnci/djr483>
- Dabiri, S., Huntsman, D., Makretsov, N., Cheang, M., Gilks, B., Bajdik, C., Gelmon, K., Chia,

- S., & Hayes, M. (2004). The presence of stromal mast cells identifies a subset of invasive breast cancers with a favorable prognosis. *Modern Pathology*, 17(6), 690-695. <https://doi.org/10.1038/modpathol.3800094>
- de Visser, K. E., & Joyce, J. A. (2023). The evolving tumor microenvironment: From cancer initiation to metastatic outgrowth. *Cancer Cell*, 41(3), 374-403. <https://doi.org/10.1016/j.ccell.2023.02.016>
- Del Prete, A., Salvi, V., Soriani, A., Laffranchi, M., Sozio, F., Bosisio, D., & Sozzani, S. (2023). Dendritic cell subsets in cancer immunity and tumor antigen sensing. *Cellular & Molecular Immunology*, 20(5), 432-447. <https://doi.org/10.1038/s41423-023-00990-6>
- Deng, W., Gowen, B. G., Zhang, L., Wang, L., Lau, S., Iannello, A., Xu, J., Rovis, T. L., Xiong, N., & Raulet, D. H. (2015). Antitumor immunity. A shed NKG2D ligand that promotes natural killer cell activation and tumor rejection. *Science*, 348(6230), 136-139. <https://doi.org/10.1126/science.1258867>
- Dhatchinamoorthy, K., Colbert, J. D., & Rock, K. L. (2021). Cancer Immune Evasion Through Loss of MHC Class I Antigen Presentation. *Frontiers in Immunology*, 12. <https://doi.org/10.3389/fimmu.2021.636568>
- Diefenbach, A., & Raulet, D. H. (1999). Natural killer cells: stress out, turn on, tune in. *Current Biology*, 9(22), R851-853. [https://doi.org/10.1016/s0960-9822\(00\)80044-5](https://doi.org/10.1016/s0960-9822(00)80044-5)
- Dighe, A. S., Richards, E., Old, L. J., & Schreiber, R. D. (1994). Enhanced in vivo growth and resistance to rejection of tumor cells expressing dominant negative IFN gamma receptors. *Immunity*, 1(6), 447-456. [https://doi.org/10.1016/1074-7613\(94\)90087-6](https://doi.org/10.1016/1074-7613(94)90087-6)
- DiLillo, D. J., Yanaba, K., & Tedder, T. F. (2010). B cells are required for optimal CD4+ and CD8+ T cell tumor immunity: therapeutic B cell depletion enhances B16 melanoma growth in mice. *The Journal of Immunology*, 184(7), 4006-4016. <https://doi.org/10.4049/jimmunol.0903009>
- Dixon, K. J., Wu, J., & Walcheck, B. (2021). Engineering Anti-Tumor Monoclonal Antibodies and Fc Receptors to Enhance ADCC by Human NK Cells. *Cancers (Basel)*, 13(2). <https://doi.org/10.3390/cancers13020312>
- Duan, Z., & Luo, Y. (2021). Targeting macrophages in cancer immunotherapy. *Signal Transduction and Targeted Therapy*, 6(1), 127. <https://doi.org/10.1038/s41392-021-00506-6>
- Dunn, G. P., Bruce, A. T., Ikeda, H., Old, L. J., & Schreiber, R. D. (2002^a). Cancer immunoediting: from immunosurveillance to tumor escape. *Nature Immunology*, 3(11), 991-998. <https://doi.org/10.1038/ni1102-991>
- Dunn, G. P., Old, L. J., & Schreiber, R. D. (2004^b). The immunobiology of cancer immunosurveillance and immunoediting. *Immunity*, 21(2), 137-148. <https://doi.org/10.1016/j.immuni.2004.07.017>
- Dunn, G. P., Old, L. J., & Schreiber, R. D. (2004^c). The three Es of cancer immunoediting. *Annual Review of Immunology*, 22, 329-360. <https://doi.org/10.1146/annurev.immunol.22.012703.104803>
- Dvorak, H. F. (1986). Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. *The New England Journal of Medicine*, 315(26), 1650-1659. <https://doi.org/10.1056/nejm198612253152606>
- Fearon, E. R. (1997). Human cancer syndromes: clues to the origin and nature of cancer. *Science*, 278(5340), 1043-1050. <https://doi.org/10.1126/science.278.5340.1043>
- Feng, M., Chen, J. Y., Weissman-Tsukamoto, R., Volkmer, J. P., Ho, P. Y., McKenna, K. M., Cheshier, S., Zhang, M., Guo, N., Gip, P., Mitra, S. S., & Weissman, I. L. (2015). Macrophages

- eat cancer cells using their own calreticulin as a guide: roles of TLR and Btk. *Proceedings of the National Academy of Sciences of the United States of America*, 112(7), 2145-2150. <https://doi.org/10.1073/pnas.1424907112>
- Feng, M., Jiang, W., Kim, B. Y. S., Zhang, C. C., Fu, Y.-X., & Weissman, I. L. (2019). Phagocytosis checkpoints as new targets for cancer immunotherapy. *Nature Reviews Cancer*, 19(10), 568-586. <https://doi.org/10.1038/s41568-019-0183-z>
- Fischer, J. R., Stehr, S., Lahm, H., Falk, W., Drings, P., & Krammer, P. H. (1992). Different Cytokines with Autocrine and Immunosuppressive Activities are Secreted by a Small-Cell Lung Cancer Cell Line. *Cytokines in Hemopoiesis, Oncology, and AIDS II*, Berlin, Heidelberg, Springer Berlin Heidelberg. https://doi.org/10.1007/978-3-642-48715-6_15
- Fridlender, Z. G., Sun, J., Kim, S., Kapoor, V., Cheng, G., Ling, L., Worthen, G. S., & Albelda, S. M. (2009). Polarization of tumor-associated neutrophil phenotype by TGF-beta: "N1" versus "N2" TAN. *Cancer Cell*, 16(3), 183-194. <https://doi.org/10.1016/j.ccr.2009.06.017>
- Furumaya, C., Martinez-Sanz, P., Bouti, P., Kuijpers, T. W., & Matlung, H. L. (2020). Plasticity in Pro- and Anti-tumor Activity of Neutrophils: Shifting the Balance. *Frontiers in Immunology*, 11, 2100. <https://doi.org/10.3389/fimmu.2020.02100>
- Galluzzi, L., Vitale, I., Warren, S., Adjemian, S., Agostinis, P., Martinez, A. B., Chan, T. A., Coukos, G., Demaria, S., Deutsch, E., Draganov, D., Edelson, R. L., Formenti, S. C., Fucikova, J., Gabriele, L., Gaip, U. S., Gameiro, S. R., Garg, A. D., Golden, E.,...Marincola, F. M. (2020). Consensus guidelines for the definition, detection and interpretation of immunogenic cell death. *The Journal for ImmunoTherapy of Cancer*, 8(1). <https://doi.org/10.1136/jitc-2019-000337>
- Gao, J., Liang, Y., & Wang, L. (2022). Shaping Polarization Of Tumor-Associated Macrophages In Cancer Immunotherapy. *Frontiers in Immunology*, 13, 888713. <https://doi.org/10.3389/fimmu.2022.888713>
- Garutti, M., Foffano, L., Mazzeo, R., Michelotti, A., Da Ros, L., Viel, A., Miolo, G., Zambelli, A., & Puglisi, F. (2023). Hereditary Cancer Syndromes: A Comprehensive Review with a Visual Tool. *Genes (Basel)*, 14(5). <https://doi.org/10.3390/genes14051025>
- Girardi, M., Oppenheim, D. E., Steele, C. R., Lewis, J. M., Glusac, E., Filler, R., Hobby, P., Sutton, B., Tigelaar, R. E., & Hayday, A. C. (2001). Regulation of cutaneous malignancy by gammadelta T cells. *Science*, 294(5542), 605-609. <https://doi.org/10.1126/science.1063916>
- Gong, T., Liu, L., Jiang, W., & Zhou, R. (2020). DAMP-sensing receptors in sterile inflammation and inflammatory diseases. *Nature Reviews Immunology*, 20(2), 95-112. <https://doi.org/10.1038/s41577-019-0215-7>
- Gonzalez-Aparicio, M., & Alfaro, C. (2020). Significance of the IL-8 pathway for immunotherapy. *Human Vaccines & Immunotherapeutics*, 16(10), 2312-2317. <https://doi.org/10.1080/21645515.2019.1696075>
- Gross, C., Holler, E., Stangl, S., Dickinson, A., Pockley, A. G., Asea, A. A., Mallappa, N., & Multhoff, G. (2008). An Hsp70 peptide initiates NK cell killing of leukemic blasts after stem cell transplantation. *Leukemia Research*, 32(4), 527-534. <https://doi.org/10.1016/j.leukres.2007.03.027>
- Gunderson, A. J., & Coussens, L. M. (2013). B cells and their mediators as targets for therapy in solid tumors. *Experimental Cell Research*, 319(11), 1644-1649. <https://doi.org/10.1016/j.yexcr.2013.03.005>

- Hamdan, F., & Cerullo, V. (2023). Cancer immunotherapies: A hope for the incurable? *Frontiers in Molecular Medicine*, 3. <https://doi.org/10.3389/fmmed.2023.1140977>
- Hanahan, D. (2022). Hallmarks of Cancer: New Dimensions. *Cancer Discovery*, 12(1), 31-46. <https://doi.org/10.1158/2159-8290.cd-21-1059>
- Hanahan, D., & Weinberg, R. A. (2000). The hallmarks of cancer. *Cell*, 100(1), 57-70. [https://doi.org/10.1016/s0092-8674\(00\)81683-9](https://doi.org/10.1016/s0092-8674(00)81683-9)
- Hellström, I., Hellstrom, K. E., Pierce, G. E., & Yang, J. P. (1968). Cellular and humoral immunity to different types of human neoplasms. *Nature*, 220(5174), 1352-1354. <https://doi.org/10.1038/2201352a0>
- Herberman, R. B., & Holden, H. T. (1978). Natural cell-mediated immunity. *Advances in Cancer Research*, 27, 305-377. [https://doi.org/10.1016/s0065-230x\(08\)60936-7](https://doi.org/10.1016/s0065-230x(08)60936-7)
- Ho, W. W., Pittet, M. J., Fukumura, D., & Jain, R. K. (2022). The local microenvironment matters in preclinical basic and translational studies of cancer immunology and immunotherapy. *Cancer Cell*, 40(7), 701-702. <https://doi.org/10.1016/j.ccell.2022.05.016>
- Jablonska, J., Lang, S., Sionov, R. V., & Granot, Z. (2017). The regulation of pre-metastatic niche formation by neutrophils. *Oncotarget*, 8(67), 112132-112144. <https://doi.org/10.18632/oncotarget.22792>
- Jahn, A., Rump, A., Widmann, T. J., Heining, C., Horak, P., Hutter, B., Paramasivam, N., Uhrig, S., Gieldon, L., Drukewitz, S., Kubler, A., Bermudez, M., Hackmann, K., Porrmann, J., Wagner, J., Arlt, M., Franke, M., Fischer, J., Kowalzyk, Z.,...Klink, B. (2022). Comprehensive cancer predisposition testing within the prospective MASTER trial identifies hereditary cancer patients and supports treatment decisions for rare cancers. *Annals of Oncology*, 33(11), 1186-1199. <https://doi.org/10.1016/j.annonc.2022.07.008>
- Janjic, B. M., Kulkarni, A., Ferris, R. L., Vujanovic, L., & Vujanovic, N. L. (2022). Human B Cells Mediate Innate Anti-Cancer Cytotoxicity Through Concurrent Engagement of Multiple TNF Superfamily Ligands. *Frontiers in Immunology*, 13. <https://doi.org/10.3389/fimmu.2022.837842>
- Jungbluth, A. A., Chen, Y.-T., Stockert, E., Busam, K. J., Kolb, D., Iversen, K., Coplan, K., Williamson, B., Altorki, N., & Old, L. J. (2001). Immunohistochemical analysis of NY-ESO-1 antigen expression in normal and malignant human tissues. *International Journal of Cancer*, 92(6), 856-860. <https://doi.org/10.1002/ijc.1282v>
- Kaplan, D. H., Shankaran, V., Dighe, A. S., Stockert, E., Aguet, M., Old, L.J., & Schreiber, R. D. (1998). Demonstration of an interferon gamma-dependent tumor surveillance system in immunocompetent mice. *Proceedings of the National Academy of Sciences of the United States of America*, 95(13), 7556-7561. <https://doi.org/10.1073/pnas.95.13.7556>
- Karre, K. (1997). How to recognize a foreign submarine. *Immunological Reviews*, 155, 5-9. <https://doi.org/10.1111/j.1600-065x.1997.tb00935.x>
- Knuth, A., Danowski, B., Oettgen, H. F., & Old, L. J. (1984). T-cell-mediated cytotoxicity against autologous malignant melanoma: analysis with interleukin 2-dependent T-cell cultures. *Proceedings of the National Academy of Sciences of the United States of America*, 81(11), 3511-3515. <https://doi.org/10.1073/pnas.81.11.3511>
- Koh, C. H., Lee, S., Kwak, M., Kim, B. S., & Chung, Y. (2023). CD8 T-cell subsets: heterogeneity, functions, and therapeutic potential. *Experimental & Molecular Medicine*, 55(11), 2287-2299. <https://doi.org/10.1038/s12276-023-01105-x>

- Kolev, M., Das, M., Gerber, M., Baver, S., Deschatelets, P., & Markiewski, M. M. (2022). Inside-Out of Complement in Cancer. *Frontiers in Immunology*, 13, 931273. <https://doi.org/10.3389/fimmu.2022.931273>
- Korman, A. J., Peggs, K. S., & Allison, J. P. (2006). Checkpoint blockade in cancer immunotherapy. *Advances in Immunology*, 90, 297-339. [https://doi.org/10.1016/s0065-2776\(06\)90008-x](https://doi.org/10.1016/s0065-2776(06)90008-x)
- Kusakabe, M., Taguchi, A., Sone, K., Mori, M., & Osuga, Y. (2023). Carcinogenesis and management of human papillomavirus-associated cervical cancer. *International Journal Clinical Oncology*, 28(8), 965-974. <https://doi.org/10.1007/s10147-023-02337-7>
- Lambert, A. W., Pattabiraman, D. R., & Weinberg, R. A. (2017). Emerging Biological Principles of Metastasis. *Cell*, 168(4), 670-691. <https://doi.org/10.1016/j.cell.2016.11.037>
- Lanier, L. L. (2008). Up on the tightrope: natural killer cell activation and inhibition. *Nature Immunology*, 9(5), 495-502. <https://doi.org/10.1038/ni1581>
- Lazar-Molnar, E., Hegyesi, H., Toth, S., & Falus, A. (2000). Autocrine and paracrine regulation by cytokines and growth factors in melanoma. *Cytokine*, 12(6), 547-554. <https://doi.org/10.1006/cyto.1999.0614>
- LeBien, T. W., & Tedder, T. F. (2008). B lymphocytes: how they develop and function. *Blood*, 112(5), 1570-1580. <https://doi.org/10.1182/blood-2008-02-078071>
- Lichtenstein, P., Holm, N. V., Verkasalo, P. K., Iliadou, A., Kaprio, J., Koskenvuo, M., Pukkala, E., Skytthe, A., & Hemminki, K. (2000). Environmental and heritable factors in the causation of cancer—analyses of cohorts of twins from Sweden, Denmark, and Finland. *N The New England Journal of Medicine*, 343(2), 78-85. <https://doi.org/10.1056/nejm200007133430201>
- Liu, E., Marin, D., Banerjee, P., Macapinlac, H. A., Thompson, P., Basar, R., Kerbauy, L. N., Overman, B., Thall, P., Kaplan, M., Nandivada, V., Kaur, I., Nunez Cortes, A., Cao, K., Daher, M., Hosing, C., Cohen, E. N., Kebriaei, P., Mehta, R.,...Rezvani, K. (2020). "Use of CAR-Transduced Natural Killer Cells in CD19-Positive Lymphoid Tumors." *The New England Journal of Medicine*, 382(6), 545-553. <https://doi.org/10.1056/nejmoa1910607>
- Liu, J., Chen, Z., Li, Y., Zhao, W., Wu, J., & Zhang, Z. (2021). PD-1/PD-L1 Checkpoint Inhibitors in Tumor Immunotherapy. *Frontiers in Pharmacology*, 12. <https://doi.org/10.3389/fphar.2021.731798>
- Liu, S., Matsuzaki, J., Wei, L., Tsuji, T., Battaglia, S., Hu, Q., Cortes, E., Wong, L., Yan, L., Long, M., Miliotto, A., Bateman, N. W., Lele, S. B., Chodon, T., Koya, R. C., Yao, S., Zhu, Q., Conrads, T. P., Wang, J.,...Odunsi, K. (2019). Efficient identification of neoantigen-specific T-cell responses in advanced human ovarian cancer. *The Journal for ImmunoTherapy of Cancer*, 7(1), 156. <https://doi.org/10.1186/s40425-019-0629-6>
- Mantovani, A., Biswas, S. K., Galdiero, M. R., Sica, A., & Locati, M. (2013). Macrophage plasticity and polarization in tissue repair and remodelling. *The Journal of Pathology*, 229(2), 176-185. <https://doi.org/10.1002/path.4133>
- Matzinger, P. (1994). Tolerance, Danger, and the Extended Family. *Annual Review of Immunology*, 12(1), 991-1045. <https://doi.org/10.1146/annurev.iy.12.040194.005015>
- Mauri, C., & Bosma, A. (2012). Immune regulatory function of B cells. *Annual Review of Immunology*, 30, 221-241. <https://doi.org/10.1146/annurev-immunol-020711-074934>
- Melillo, R. M., Guarino, V., Avilla, E., Galdiero, M. R., Liotti, F., Prevete, N., Rossi, F. W., Basolo, F., Ugolini, C., de Paulis, A., Santoro, M., & Marone, G. (2010). Mast cells have a protumorigenic role in human thyroid cancer. *Oncogene*, 29(47), 6203-6215. <https://doi.org/10.1038/ng.6203>

- [org/10.1038/onc.2010.348](https://doi.org/10.1038/onc.2010.348)
- Mills, C. D., Kincaid, K., Alt, J. M., Heilman, M. J., & Hill, A. M. (2000). M-1/M-2 macrophages and the Th1/Th2 paradigm. *The Journal of Immunology*, 164(12), 6166-6173. <https://doi.org/10.4049/jimmunol.164.12.6166>
- Mills, C. D., & Ley, K. (2014). M1 and M2 macrophages: the chicken and the egg of immunity. *Journal of Innate Immunity*, 6(6), 716-726. <https://doi.org/10.1159/000364945>
- Neale, R. E., Lucas, R. M., Byrne, S. N., Hollestein, L., Rhodes, L. E., Yazar, S., Young, A. R., Berwick, M., Ireland, R. A., & Olsen, C. M. (2023). The effects of exposure to solar radiation on human health. *Photochemical & Photobiological Sciences*, 22(5), 1011-1047. <https://doi.org/10.1007/s43630-023-00375-8>
- Nersesian, S., Carter, E. B., Lee, S. N., Westhaver, L. P., & Boudreau, J. E. (2023). Killer instincts: natural killer cells as multifactorial cancer immunotherapy. *Frontiers in Immunology*, 14, 1269614. <https://doi.org/10.3389/fimmu.2023.1269614>
- Nishizuka, Y., Nakakuki, K., & Usui, M. (1965). Enhancing Effect of Thymectomy on Hepatotumorigenesis in Swiss Mice following Neonatal Injection of 20-Methylcholanthrene. *Nature*, 205(4977), 1236-1238. <https://doi.org/10.1038/2051236b0>
- Noessner, E., Gastpar, R., Milani, V., Brandl, A., Hutzler, P. J., Kuppner, M. C., Roos, M., Kremmer, E., Asea, A., Calderwood, S. K., & Issels, R. D. (2002). Tumor-derived heat shock protein 70 peptide complexes are cross-presented by human dendritic cells. *Journal of Immunology*, 169(10), 5424-5432. <https://doi.org/10.4049/jimmunol.169.10.5424>
- Nussenzweig, M. C., Steinman, R. M., Gutchinov, B., & Cohn, Z. A. (1980). Dendritic cells are accessory cells for the development of anti-trinitrophenyl cytotoxic T lymphocytes. *Journal of Experimental Medicine*, 152(4), 1070-1084. <https://doi.org/10.1084/jem.152.4.1070>
- Paijens, S. T., Vledder, A., de Bruyn, M., & Nijman, H. W. (2021). Tumor-infiltrating lymphocytes in the immunotherapy era. *Cellular & Molecular Immunology*, 18(4), 842-859. <https://doi.org/10.1038/s41423-020-00565-9>
- Plesca, I., Muller, L., Bottcher, J. P., Medyouf, H., Wehner, R., & Schmitz, M. (2022). Tumor-associated human dendritic cell subsets: Phenotype, functional orientation, and clinical relevance. *European Journal of Immunology*, 52(11), 1750-1758. <https://doi.org/10.1002/eji.202149487>
- Quezada, S. A., Simpson, T. R., Peggs, K. S., Merghoub, T., Vider, J., Fan, X., Blasberg, R., Yagita, H., Muranski, P., Antony, P. A., Restifo, N. P., & Allison, J. P. (2010). Tumor-reactive CD4(+) T cells develop cytotoxic activity and eradicate large established melanoma after transfer into lymphopenic hosts. *Journal of Experimental Medicine*, 207(3), 637-650. <https://doi.org/10.1084/jem.20091918>
- Rabenhorst, A., Schlaak, M., Heukamp, L. C., Forster, A., Theurich, S., von Bergwelt-Baildon, M., Buttner, R., Kurschat, P., Mauch, C., Roers, A., & Hartmann, K. (2012). Mast cells play a protumorigenic role in primary cutaneous lymphoma. *Blood*, 120(10), 2042-2054. <https://doi.org/10.1182/blood-2012-03-415638>
- Rakae, M., Busund, L. T., Paulsen, E. E., Richardsen, E., Al-Saad, S., Andersen, S., Donnem, T., Bremnes, R. M., & Kilvaer, T. K. (2016). Prognostic effect of intratumoral neutrophils across histological subtypes of non-small cell lung cancer. *Oncotarget*, 7(44), 72184-72196. <https://doi.org/10.18632/oncotarget.12360>
- Rao, H. L., Chen, J. W., Li, M., Xiao, Y. B., Fu, J., Zeng, Y. X., Cai, M. Y., & Xie, D. (2012).

- Increased intratumoral neutrophil in colorectal carcinomas correlates closely with malignant phenotype and predicts patients' adverse prognosis. *PLoS One*, 7(1), e30806. <https://doi.org/10.1371/journal.pone.0030806>
- Rao, Q., Chen, Y., Yeh, C. R., Ding, J., Li, L., Chang, C., & Yeh, S. (2016). Recruited mast cells in the tumor microenvironment enhance bladder cancer metastasis via modulation of ERbeta/CCL2/CCR2 EMT/MMP9 signals. *Oncotarget*, 7(7), 7842-7855. <https://doi.org/10.18632/oncotarget.5467>
- Ravetch, J. V., & Bolland, S. (2001). IgG Fc Receptors. *Annual Review of Immunology*, 19(1), 275-290. <https://doi.org/10.1146/annurev.immunol.19.1.275>
- Reizis, B. (2019). Plasmacytoid Dendritic Cells: Development, Regulation, and Function. *Immunity*, 50(1), 37-50. <https://doi.org/10.1016/j.immuni.2018.12.027>
- Robbins, P. F., Lu, Y.-C., El-Gamil, M., Li, Y. F., Gross, C., Gartner, J., Lin, J. C., Teer, J. K., Cliften, P., Tycksen, E., Samuels, Y., & Rosenberg, S. A. (2013). Mining exomic sequencing data to identify mutated antigens recognized by adoptively transferred tumor-reactive T cells. *Nature Medicine*, 19(6), 747-752. <https://doi.org/10.1038/nm.3161>
- Sagiv, J. Y., Michaeli, J., Assi, S., Mishalian, I., Kisos, H., Levy, L., Damti, P., Lumbroso, D., Polyansky, L., Sionov, R. V., Ariel, A., Hovav, A. H., Henke, E., Fridlender, Z. G., & Granot, Z. (2015). Phenotypic diversity and plasticity in circulating neutrophil subpopulations in cancer. *Cell Reports*, 10(4), 562-573. <https://doi.org/10.1016/j.celrep.2014.12.039>
- Sahin, U., Türeci, O., Schmitt, H., Cochlovius, B., Johannes, T., Schmits, R., Stenner, F., Luo, G., Schobert, I., & Pfreundschuh, M. (1995). Human neoplasms elicit multiple specific immune responses in the autologous host. *Proceedings of the National Academy of Sciences of the United States of America*, 92(25), 11810-11813. <https://doi.org/10.1073/pnas.92.25.11810>
- Sanford, B. H., Kohn, H. I., Daly, J. J., & Soo, S. F. (1973). Long-term spontaneous tumor incidence in neonatally thymectomized mice. *Journal of Immunology*, 110(5), 1437-1439. <https://doi.org/10.4049/jimmunol.110.5.1437>
- Satie, A. P., Rajpert-De Meyts, E., Spagnoli, G. C., Henno, S., Olivo, L., Jacobsen, G. K., Rioux-Leclercq, N., Jegou, B., & Samson, M. (2002). The cancer-testis gene, NY-ESO-1, is expressed in normal fetal and adult testes and in spermatocytic seminomas and testicular carcinoma in situ. *Laboratory Investigation*, 82(6), 775-780. <https://doi.org/10.1097/01.lab.0000017169.26718.5f>
- Schlosser, H. A., Thelen, M., Lechner, A., Wennhold, K., Garcia-Marquez, M. A., Rothschild, S. I., Staib, E., Zander, T., Beutner, D., Gathof, B., Gilles, R., Cukuroglu, E., Goke, J., Shimabukuro-Vornhagen, A., Drebber, U., Quaas, A., Bruns, C. J., Holscher, A. H., & Von Bergwelt-Baildon, M. S. (2019). B cells in esophago-gastric adenocarcinoma are highly differentiated, organize in tertiary lymphoid structures and produce tumor-specific antibodies. *Oncoimmunology*, 8(1), e1512458. <https://doi.org/10.1080/2162402x.2018.1512458>
- Schreiber, R. D., Old, L. J., & Smyth, M. J. (2011). Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. *Science*, 331(6024), 1565-1570. <https://doi.org/10.1126/science.1203486>
- See, P., Dutertre, C. A., Chen, J., Gunther, P., McGovern, N., Irac, S. E., Gunawan, M., Beyer, M., Handler, K., Duan, K., Sumatoh, H. R. B., Ruffin, N., Jouve, M., Gea-Mallorqui, Hennekam, E., R. C. M., Lim, T., Yip, C. C., Wen, M., Malleret, B.,... Ginhoux, F. (2017). Mapping the human DC lineage through the integration of high-dimensional techniques. *Science*, 356(6342).

- <https://doi.org/10.1126/science.aag3009>
- Shankaran, V., Ikeda, H., Bruce, A. T., White, J. M., Swanson, P. E., Old, L. J., & Schreiber, R. D. (2001). IFN γ and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature*, 410(6832), 1107-1111. <https://doi.org/10.1038/35074122>
- Shapouri-Moghaddam, A., Mohammadian, S., Vazini, H., Taghadosi, M., Esmaeili, S. A., Mardani, F., Seifi, B., Mohammadi, A., Afshari, J. T., & Sahebkar, A. (2018). Macrophage plasticity, polarization, and function in health and disease. *Journal of Cellular Physiology*, 233(9), 6425-6440. <https://doi.org/10.1002/jcp.26429>
- Shaul, M. E., & Fridlender, Z. G. (2019). Tumour-associated neutrophils in patients with cancer. *Nature Reviews Clinical Oncology*, 16(10), 601-620. <https://doi.org/10.1038/s41571-019-0222-4>
- Siegel, F. P., Kadowaki, N., Shodell, M., Fitzgerald-Bocarsly, P. A., Shah, K., Ho, S., Antonenko, S., & Liu, Y. J. (1999). The nature of the principal type 1 interferon-producing cells in human blood. *Science*, 284(5421), 1835-1837. <https://doi.org/10.1126/science.284.5421.1835>
- Smith, C. C., Selitsky, S. R., Chai, S., Armistead, P. M., Vincent, B. G., & Serody, J. S. (2019). Alternative tumour-specific antigens. *Nature Reviews Cancer*, 19(8), 465-478. <https://doi.org/10.1038/s41568-019-0162-4>
- Smyth, M. J., Dunn, G. P., & Schreiber, R. D. (2006^a). Cancer immunosurveillance and immunoediting: the roles of immunity in suppressing tumor development and shaping tumor immunogenicity. *Advances in Immunology*, 90, 1-50. [https://doi.org/10.1016/s0065-2776\(06\)90001-7](https://doi.org/10.1016/s0065-2776(06)90001-7)
- Smyth, M. J., Thia, K. Y., Street, S. E., Cretney, E., Trapani, J. A., Taniguchi, M., Kawano, T., Pelikan, S. B., Crowe, N. Y., & Godfrey, D. I. (2000^b). Differential tumor surveillance by natural killer (NK) and NKT cells. *Journal of Experimental Medicine*, 191(4), 661-668. <https://doi.org/10.1084/jem.191.4.661>
- Smyth, M. J., Thia, K. Y., Street, S. E., MacGregor, D., Godfrey, D. I., & Trapani, J. A. (2000^c). Perforin-mediated cytotoxicity is critical for surveillance of spontaneous lymphoma. *Journal of Experimental Medicine*, 192(5), 755-760. <https://doi.org/10.1084/jem.191.4.661>
- Sondel, P. M., & Hank, J. A. (2001). Antibody-directed, effector cell-mediated tumor destruction. *Hematology/Oncology Clinics of North America*, 15(4), 703-721. [https://doi.org/10.1016/s0889-8588\(05\)70243-4](https://doi.org/10.1016/s0889-8588(05)70243-4)
- Steinman, R. M. (2012). Decisions about dendritic cells: past, present, and future. *Annual Reviews Immunology*, 30, 1-22. <https://doi.org/10.1146/annurev-immunol-100311-102839>
- Street, S. E., Trapani, J. A., MacGregor, D., & Smyth, M. J. (2002). Suppression of lymphoma and epithelial malignancies effected by interferon gamma. *Journal of Experimental Medicine*, 196(1), 129-134. <https://doi.org/10.1084/jem.20020063>
- Stutman, O. (1975). Immunodepression and malignancy. *Advances in Cancer Research*, 22, 261-422. [https://doi.org/10.1016/s0065-230x\(08\)60179-7](https://doi.org/10.1016/s0065-230x(08)60179-7)
- Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., & Bray, F. (2021). Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA: A Cancer Journal for Clinicians*, 71(3), 209-249. <https://doi.org/10.3322/caac.21660>
- Sykulev, Y., Joo, M., Vturina, I., Tsomides, T. J., & Eisen, H. N. (1996). Evidence that a single peptide-MHC complex on a target cell can elicit a cytolytic T cell response. *Immunity*, 4(6),

- 565-571. [https://doi.org/10.1016/s1074-7613\(00\)80483-5](https://doi.org/10.1016/s1074-7613(00)80483-5)
- Tan, R., Nie, M., & Long, W. (2022). The role of B cells in cancer development. *Frontiers in Oncology*, 12, 958756. <https://doi.org/10.3389/fonc.2022.958756>
- Thomas, R., Al-Khadairi, G., Roelands, J., Hendrickx, W., Dermime, S., Bedognetti, D., & Decock, J. (2018). NY-ESO-1 Based Immunotherapy of Cancer: Current Perspectives. *Frontiers in Immunology*, 9, 947. <https://doi.org/10.3389/fimmu.2018.00947>
- Titus, J. A., Perez, P., Kaubisch, A., Garrido, M. A., & Segal, D. M. (1987). Human K/natural killer cells targeted with hetero-cross-linked antibodies specifically lyse tumor cells in vitro and prevent tumor growth in vivo. *Journal in Immunology*, 139(9), 3153-3158. <https://doi.org/10.4049/jimmunol.139.9.3153>
- Togashi, Y., Shitara, K., & Nishikawa, H. (2019). Regulatory T cells in cancer immunosuppression — implications for anticancer therapy. *Nature Reviews Clinical Oncology*, 16(6), 356-371. <https://doi.org/10.1038/s41571-019-0175-7>
- Tomasetti, C., & Vogelstein, B. (2015). Cancer etiology. Variation in cancer risk among tissues can be explained by the number of stem cell divisions. *Science*, 347(6217), 78-81. <https://doi.org/10.1126/science.1260825>
- Vallabhapurapu, S. D., Blanco, V. M., Sulaiman, M. K., Vallabhapurapu, S. L., Chu, Z., Franco, R. S., & Qi, X. (2015). Variation in human cancer cell external phosphatidylserine is regulated by flippase activity and intracellular calcium. *Oncotarget*, 6(33), 34375-34388. <https://doi.org/10.18632/oncotarget.6045>
- van der Bruggen, P., Traversari, C., Chomez, P., Lurquin, C., De Plaen, E., Van den Eynde, B., Knuth, A., & Boon, T. (1991). A gene encoding an antigen recognized by cytolytic T lymphocytes on a human melanoma. *Science*, 254(5038), 1643-1647. <http://doi.org/10.1126/science.1840703>
- van Leeuwen, E. M., Remmerswaal, E. B., Vossen, M. T., Rowshani, A. T., Wertheim-van Dillen, P. M., van Lier, R. A., & ten Berge, I. J. (2004). Emergence of a CD4+CD28- granzyme B+, cytomegalovirus-specific T cell subset after recovery of primary cytomegalovirus infection. *Journal in Immunology*, 173(3), 1834-1841. <https://doi.org/10.4049/jimmunol.173.3.1834>
- Varricchi, G., Galdiero, M. R., Loffredo, S., Marone, G., Iannone, R., Marone, G., & Granata, F. (2017). Are Mast Cells MASTers in Cancer? *Frontiers in Immunology*, 8, 424. <https://doi.org/10.3389/fimmu.2017.00424>
- Vesely, M. D., Kershaw, M. H., Schreiber, R. D., & Smyth, M. J. (2011). Natural innate and adaptive immunity to cancer. *Annual Reviews of Immunology*, 29, 235-271. <https://doi.org/10.1146/annurev-immunol-031210-101324>
- West, W. H., Cannon, G. B., Kay, H. D., Bonnard, G. D., & Herberman, R. B. (1977). Natural Cytotoxic Reactivity of Human Lymphocytes Against a Myeloid Cell Line: Characterization of Effector Cells. *The Journal of Immunology*, 118(1), 355-361. <https://doi.org/10.4049/jimmunol.118.1.355>
- Wherry, E. J. (2011). T cell exhaustion. *Nature Immunology*, 12(6), 492-499. <https://doi.org/10.1038/ni.2035>
- White, E., Kirkpatrick, C. S., & Lee, J. A. (1994). Case-control study of malignant melanoma in Washington State. I. Constitutional factors and sun exposure. *American Journal of Epidemiology*, 139(9), 857-868. <https://doi.org/10.1093/oxfordjournals.aje.a117092>
- Woloszynska-Read, A., Mhaweche-Fauceglia, P., Yu, J., Odunsi, K., & Karpf, A. R. (2008).

- Intertumor and intratumor NY-ESO-1 expression heterogeneity is associated with promoter-specific and global DNA methylation status in ovarian cancer. *Clinical Cancer Research*, 14(11), 3283-3290. <https://doi.org/10.1158/1078-0432.ccr-07-5279>
- Wu, S., Powers, S., Zhu, W., & Hannun, Y.A. (2016). Substantial contribution of extrinsic risk factors to cancer development. *Nature*, 529(7584), 43-47. <https://doi.org/10.1038/nature16166>
- Wu, S., Zhu, W., Thompson, P., & Hannun, Y. A. (2018). Evaluating intrinsic and non-intrinsic cancer risk factors. *Nature Communications*, 9(1), 3490. <https://doi.org/10.1038/s41467-018-05467-z>
- Xing, S., & Ferrari de Andrade, L. (2020). NKG2D and MICA/B shedding: a 'tag game' between NK cells and malignant cells. *Clinical & Translational Immunology*, 9(12), e1230. <https://doi.org/10.1002/cti2.1230>
- Yamamoto, T. N., Kishton, R. J., & Restifo, N. P. (2019). Developing neoantigen-targeted T cell-based treatments for solid tumors. *Nature Medicine*, 25(10), 1488-1499. <https://doi.org/10.1038/s41591-019-0596-y>
- Yano, H., Kinuta, M., Tateishi, H., Nakano, Y., Matsui, S., Monden, T., Okamura, J., Sakai, M., & Okamoto, S. (1999). Mast cell infiltration around gastric cancer cells correlates with tumor angiogenesis and metastasis. *Gastric Cancer*, 2(1), 26-32. <https://doi.org/10.1038/sj.bjc.6603962>
- Yarchoan, M., Johnson, B. A. 3rd, Lutz, E. R., Laheru, D. A., & Jaffee, E. M. (2017). Targeting neoantigens to augment antitumour immunity. *Nature Reviews Cancer*, 17(4), 209-222. <https://doi.org/10.1038/nrc.2017.74>